

RESEARCH ARTICLE

Open Access



Notch signalling defines dorsal root ganglia neuroglial fate choice during early neural crest cell migration

Sophie Wiszniak and Quenten Schwarz*

Abstract

Background: The dorsal root ganglia (DRG) are a critical component of the peripheral nervous system, and function to relay somatosensory information from the body's periphery to sensory perception centres within the brain. The DRG are primarily comprised of two cell types, sensory neurons and glia, both of which are neural crest-derived. Notch signalling is known to play an essential role in defining the neuronal or glial fate of bipotent neural crest progenitors that migrate from the dorsal ridge of the neural tube to the sites of the DRG. However, the involvement of Notch ligands in this process and the timing at which neuronal versus glial fate is acquired has remained uncertain.

Results: We have used tissue specific knockout of the E3 ubiquitin ligase mindbomb1 (*Mib1*) to remove the function of all Notch ligands in neural crest cells. *Wnt1-Cre; Mib1^{fl/fl}* mice exhibit severe DRG defects, including a reduction in glial cells, and neuronal cell death later in development. By comparing formation of sensory neurons and glia with the expression and activation of Notch signalling in these mice, we define a critical period during embryonic development in which early migrating neural crest cells become biased toward neuronal and glial phenotypes.

Conclusions: We demonstrate active Notch signalling between neural crest progenitors as soon as trunk neural crest cells delaminate from the neural tube and during their early migration toward the site of the DRG. This data brings into question the timing of neuroglial fate specification in the DRG and suggest that it may occur much earlier than originally considered.

Keywords: Notch signalling, *Mib1*, Neural crest, Fate restriction, Dorsal root ganglia

Background

Peripheral sensory neurons and their supporting glia condense into specialised dorsal root ganglia (DRG) that transmit somatosensory information from the body's periphery to sensory perception centres within the central nervous system. A diverse array of neuronal sub-types underlies the ability of the DRG to transduce different sensations including pain, proprioception, temperature and touch. Satellite glia sit in close association with neuronal cell bodies of the sensory ganglia to modulate their microenvironment [1]. Both cell types comprising the DRG arise from a sub-population

of bipotent trunk neural crest cell progenitors that are biased towards the sensory lineage [2]. This population of trunk neural crest cells migrates ventrally from the dorsal neural tube, arrests within the somites, and is then thought to gain neuronal or glial identity as they coalesce into the DRG [3, 4].

Notch signalling is known to play an important role in neuroglial fate choice determination [5, 6]. The Notch signalling pathway relies on direct cell–cell interactions mediated by Notch receptors on the signal-receiving cell and ligands such as Delta and Jagged on the signal-sending cell. By a process known as lateral inhibition, Notch signalling enables a population of homogeneous progenitors to become specified into different cell types, namely neurons and glia in the developing nervous system. In the DRG, cells expressing Delta-like 1 (*DLL1*) adopt a

*Correspondence: quenten.schwarz@unisa.edu.au
Centre for Cancer Biology, University of South Australia and SA Pathology,
North Terrace, Adelaide, SA 5001, Australia



neuronal fate, which then signal to adjacent cells via their Notch receptors to inhibit neuronal differentiation. This in turn promotes glial differentiation of cells in which neuronal differentiation has been inhibited [7, 8].

The study of Notch signalling in DRG development has been complicated by the presence of multiple Notch receptors and ligands, all of which have the potential to act in a functionally redundant manner. This has been overcome to some degree by the study of recombination signal binding protein for immunoglobulin kappa J region (*Rbpj*) conditional knockout mice. RBPJ is a transcription factor which interacts with the intracellular domain of all Notch receptors and is essential to mediate downstream transcriptional effects upon Notch pathway stimulation. Therefore, knockout of *Rbpj* is expected to abolish all Notch signalling. Removal of *Rbpj* specifically in neural crest cells leads to profound DRG defects, including a significant reduction in glial cells [9, 10], which is consistent with a role for Notch signalling in promoting glial cell development. However, these studies did not define the timing of Notch activation during DRG development and gliogenesis, as well as the roles for Notch ligands in this process.

While several Notch ligands have been ubiquitously removed during mouse development (e.g. *Dll1-null*) their specific effects on DRG formation have been difficult to study given the broad roles Notch ligands play in many organ systems during embryonic development, and the severity of knockout phenotypes [11]. Also, the removal of individual Notch ligands may be phenotypically misleading if multiple ligands are able to function in a redundant fashion. To overcome these difficulties, we addressed the function of Notch ligands during neural crest and DRG development by removing *Mind bomb 1* (*Mib1*) conditionally in neural crest-derived tissue. Mib1 is an E3 ubiquitin ligase that is required in signal-sending cells to target the Notch ligands for ubiquitination. This ubiquitination is required for the internalisation of Notch ligands upon signalling to Notch receptors on adjacent cells, and this internalisation is essential for the correct cleavage and translocation of the Notch intracellular domain to the nucleus in the signal-receiving cell to enable active Notch signalling [12]. Therefore, removal of *Mib1* in neural crest cells is expected to abolish activity of all Notch ligands.

Consistent with the known role for Notch signalling in DRG development, *Wnt1-Cre; Mib1^{fl/fl}* embryos exhibit severe DRG hypoplasia, with a dramatic reduction in glial cells in the DRG. Interestingly, the loss of glial cells was preceded by reduced SRY-related HMG-box 10 (Sox10) expression in a subset of early migrating neural crest cells. Sox10 is a marker of pre- and migratory neural crest cells, which is lost as neural crest cells

differentiate into neurons, however is maintained in neural crest progenitors that differentiate into glia, and thus is also used as a marker of mature glial cells [13, 14]. This early loss of Sox10 presents the possibility that a subset of neural crest cells are specified to become glia at the earliest stages of their migration, before neurons differentiate. In the absence of *Mib1*, DLL1 protein accumulates at the cell membrane, and enables visualisation of neuronally biased signal-sending cells attempting to undergo active Notch signalling. In *Wnt1-Cre; Mib1^{fl/fl}* embryos, aberrant DLL1 accumulation was evident as early as E9.25 immediately after neural crest cells had delaminated from the neural tube, and this was accompanied by a loss of Notch1 intracellular domain (NICD) in the nucleus of migrating neural crest cells at this stage. This is the first study to demonstrate active Notch signalling between neural crest progenitors at this early stage of trunk neural crest migration, and suggests that the signalling events controlling neuroglial fate specification in the DRG occur much earlier than originally considered.

Results

Loss of Mib1 in neural crest cells causes severe dorsal root ganglia hypoplasia

At E12.5, the dorsal root ganglia (DRG) appear as uniformly-shaped, segmented tissue structures, positioned bi-laterally adjacent to the neural tube. All neurons and glia that comprise the mature DRG are derived from neural crest cells [15]. Removal of Mib1 specifically in neural crest-derived tissue using a *Wnt1-Cre* driver and *Mib1^{fl}* allele revealed severe DRG hypoplasia ($n=3/3$; Fig. 1). Longitudinal sections through the developing neural tube and DRG at E12.5 revealed that compared to control littermates (*Wildtype* (Cre-negative) and *Wnt1-Cre; Mib1^{fl/+}*), *Wnt1-Cre; Mib1^{fl/fl}* embryos exhibited a dramatic reduction of DRG neuronal tissue, as labelled by immunostaining for the mature neuronal marker TuJ1 (Fig. 1a). Lineage tracing of neural crest-derived tissue was performed by examining expression of a *Z/EG* reporter gene, which permanently labels cells with EGFP upon expression of the *Wnt1-Cre* driver. Similar to TuJ1, EGFP immunostaining was dramatically reduced in the DRG of *Wnt1-Cre; Mib1^{fl/fl}* embryos (Fig. 1b), indicating DRG deficiency at E12.5 is caused by a loss of all neural crest-derived cellular components of the DRG.

Loss of Mib1 results in reduced glial progenitors, premature neuronal differentiation and neuronal cell death in DRG

To investigate the developmental timing of DRG deficiency, DRG formation was examined at earlier stages of development. By E10.5, all neural crest cells that will contribute to the neuronal and glial cell subtypes of the

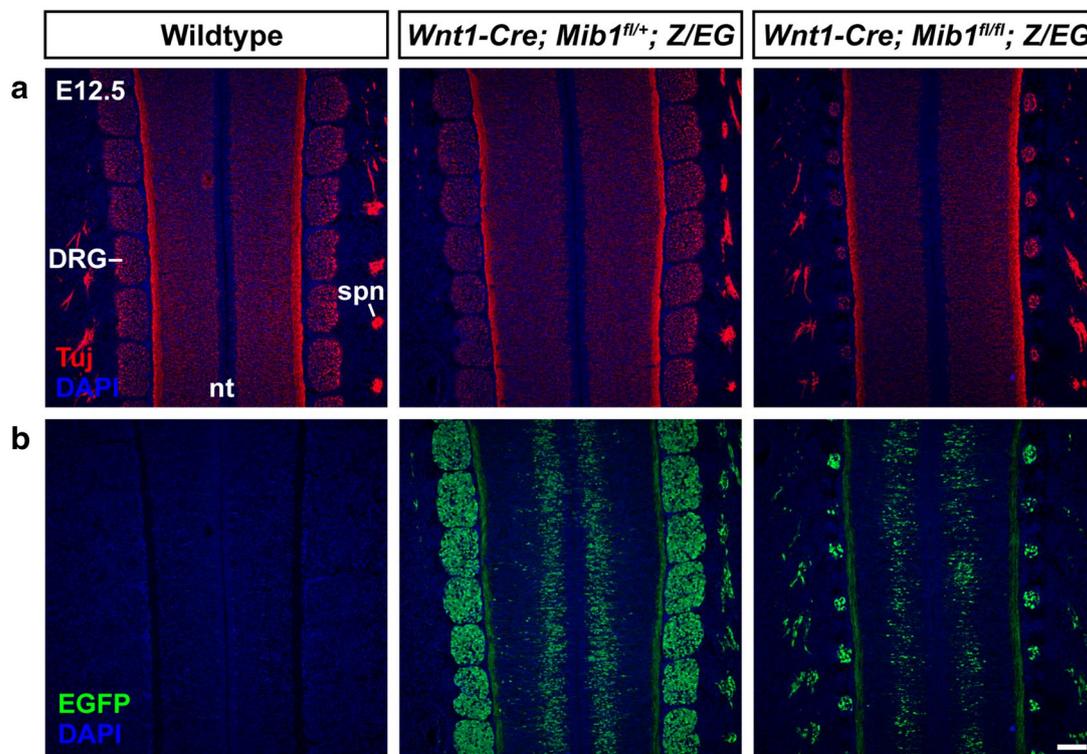


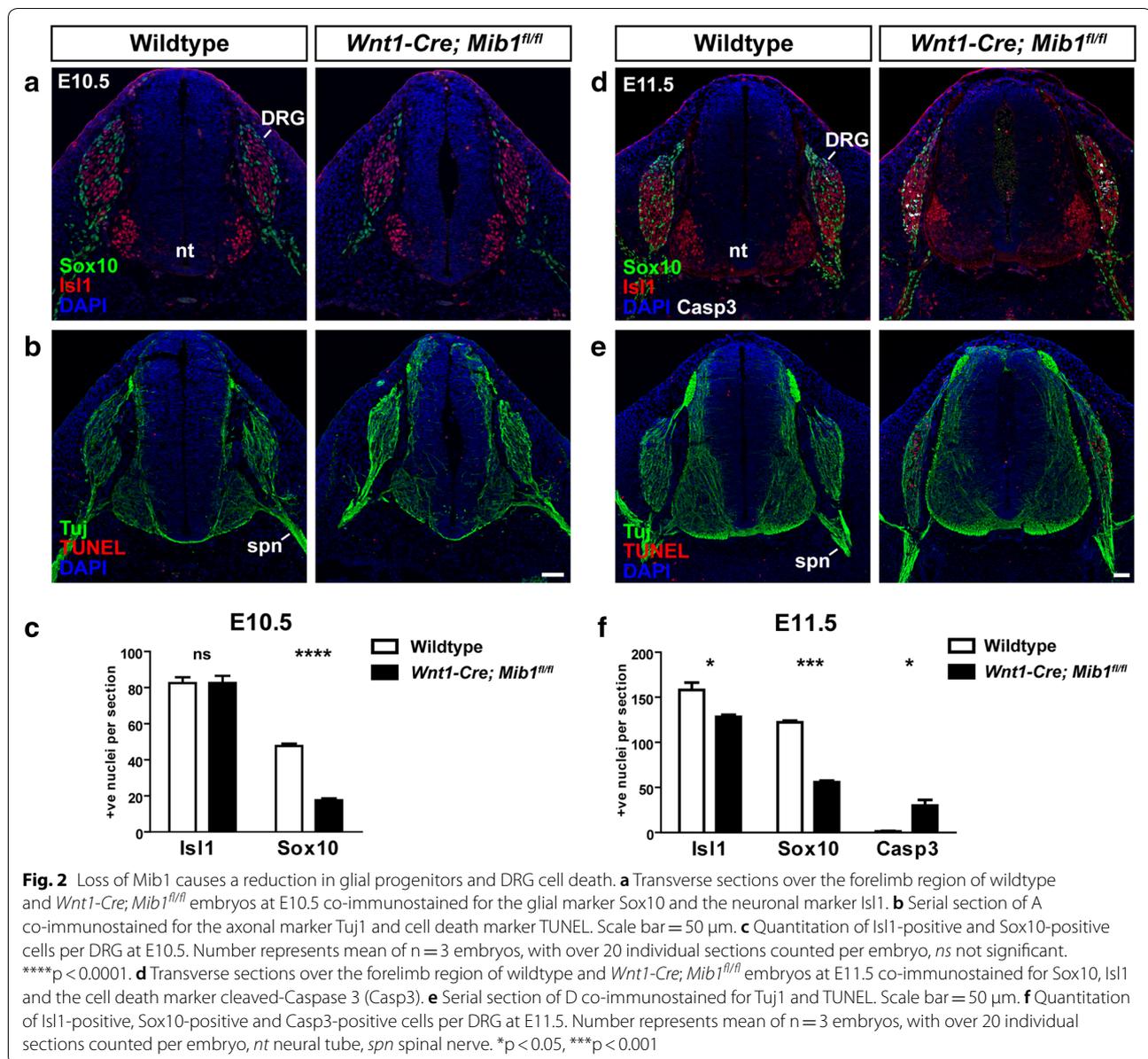
Fig. 1 Loss of Mib1 in neural crest cells causes severe DRG hypoplasia. Longitudinal sections over the forelimb region of E12.5 wildtype, *Wnt1-Cre; Mib1^{fl/fl}; Z/EG*, and *Wnt1-Cre; Mib1^{fl/fl}; Z/EG* embryos were co-immunostained for Tuj1 (a) and EGFP (b) $n = 3/3$ /genotype. Blue, DAPI. Colour channels are separated for clarity. nt, neural tube; spn, spinal nerve. Scale bar = 100 μ m

DRG have delaminated from the neural tube, and reside as a mass of cells lateral to the spinal cord. At this stage of DRG development, neuronal and glial progenitors have begun the process of specification and differentiation as defined by the markers *Isl1* for neurons and *Sox10* for glial cells. Analysis of transverse sections through the DRG over the forelimb region of E10.5 wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos revealed a dramatic reduction in *Sox10*-positive glial progenitors in *Wnt1-Cre; Mib1^{fl/fl}* embryos, without a change in the number of *Isl1*-positive neurons ($n = 3/3$; Fig. 2a, c). Immunostaining of serial sections with the neuronal marker Tuj1, further showed that neuronal differentiation proceeded normally in the DRG at E10.5 (Fig. 2b). No changes in cell death, assessed by Terminal deoxynucleotidyl transferase dUTP nick end labelling (TUNEL), were observed that may explain the loss of *Sox10*-positive glial progenitors (Fig. 2b).

By E11.5, the number of *Sox10*-positive glia and *Isl1*-positive neurons was significantly reduced in *Wnt1-Cre; Mib1^{fl/fl}* embryos compared to wildtype ($n = 3/3$; Fig. 2d, f). This was accompanied by a dramatic increase in cell death in the DRG, as assessed by cleaved-caspase 3 (Casp3) and TUNEL staining (Fig. 2d–f). Analysis of the

mature glial marker Fatty acid binding protein 7 (FABP7) also showed that gliogenesis in the DRG was severely inhibited in *Wnt1-Cre; Mib1^{fl/fl}* embryos at both E11.5 and E13.5 ($n = 3/3$; Additional file 1: Fig. S1). Our data therefore suggest that the DRG hypoplasia observed in *Wnt1-Cre; Mib1^{fl/fl}* embryos from E11.5 occurs as a result of both glial deficiency and aberrant neuronal cell death. Whether this is due to a cell-autonomous role for Mib1 in neuronal cell survival, or a secondary effect due to loss of glial cells remains to be investigated.

Sensory neurons can be divided into nociceptive, mechanoreceptive and proprioceptive subtypes based on their expression profile of the neurotrophin (Trk) receptors TrkA, TrkB and TrkC [16]. During DRG morphogenesis, TrkC-positive neurons are the first to be generated, followed temporally by TrkA and TrkB, although there is also a degree of co-expression of Trk receptors within cells at early stages of differentiation [16]. To investigate whether cell death was restricted to a particular sensory neuron subtype, Trk receptor expression was analysed in *Wnt1-Cre; Mib1^{fl/fl}* embryos at E10.5 and E11.5. At E10.5, prior to neuronal cell death occurring, expression of all Trk receptors was increased in *Wnt1-Cre; Mib1^{fl/fl}* embryos compared to wildtype ($n = 3/3$; Fig. 3a–g). At

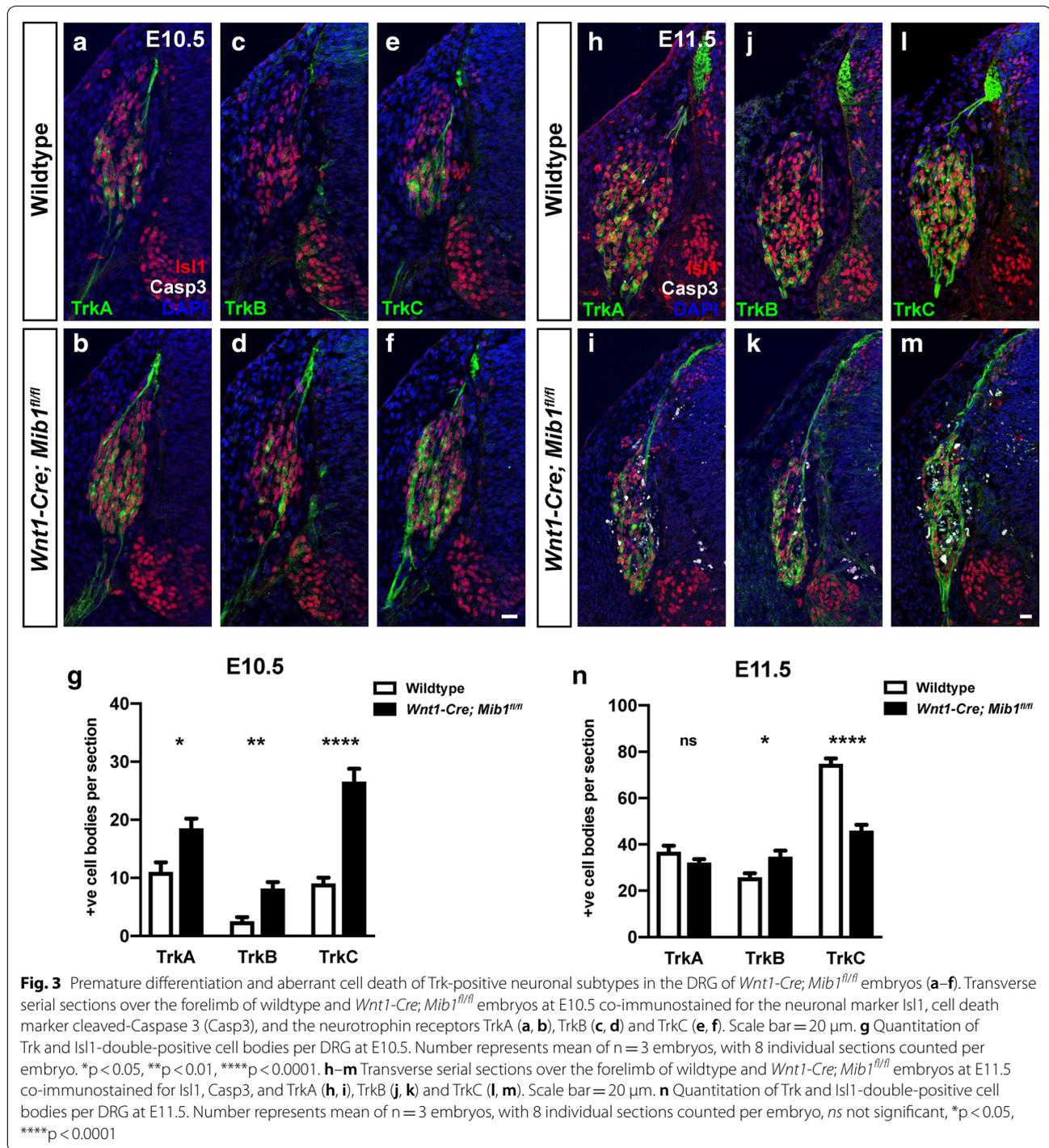


E11.5, the dramatic increase in cell death corresponded with a decrease in TrkC-positive neurons in *Wnt1-Cre; Mib1^{fl/fl}* embryos, while there was no difference in TrkA numbers, and a slight increase in TrkB-positive neurons remained (n = 3/3; Fig. 3h–n). Taken together, these results suggest initial premature differentiation of neurons occurs upon loss of Mib1, which does not appear to be subtype-specific since expression of all Trk receptors is increased. At E10.5, TrkC showed the most dramatic increase, which is consistent with the TrkC-positive population being the first to be generated. Therefore the selective loss of TrkC-positive neurons at E11.5 may reflect neuronal cell death occurring in a temporal

cascade such that the first born TrkC neurons are thus the first to die, leaving the latest born TrkB-positive neurons unaffected at this stage. Alternatively, loss of Mib1 may selectively affect survival of TrkC-positive neurons. Conditional knockout of Mib1 using alternative sensory-specific Cre-drivers may be required to further elucidate a definitive role for Mib1 in neuronal survival and in which particular neuronal subtypes.

Sox10 is downregulated rapidly during early neural crest cell migration with loss of Mib1

To further investigate the origin of reduced DRG glial progenitors present in *Wnt1-Cre; Mib1^{fl/fl}* embryos



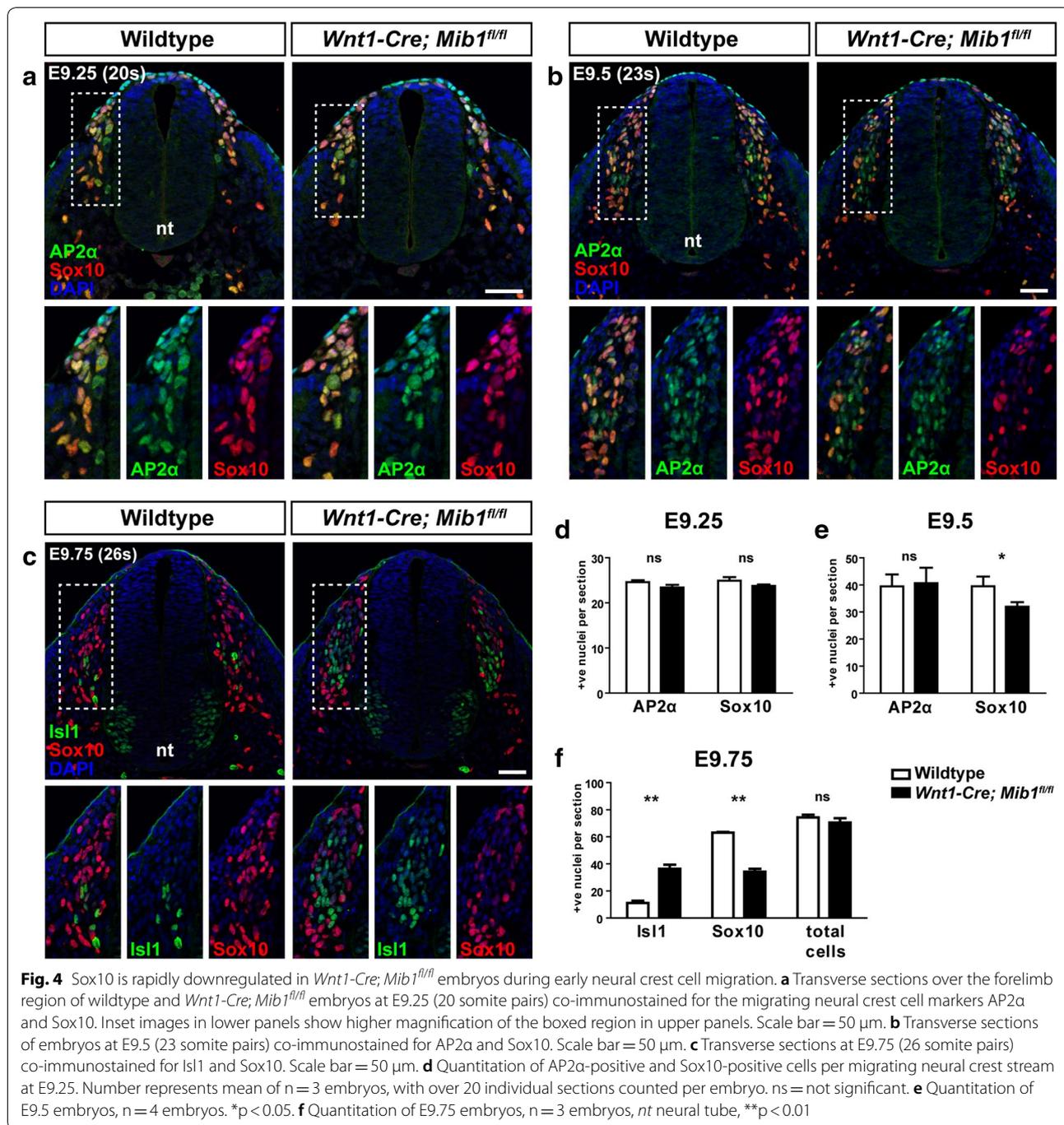
at E10.5, we investigated early stages of trunk neural crest cell development from E9.25 (20 somite) to E9.75 (26 somite). As well as a marker of glial cells, Sox10 is expressed in neural crest cells immediately upon their delamination from the neural tube, and is hence widely used as a neural crest cell marker. This has made study

of neural crest-derived glial progenitors somewhat challenging as the Sox10 marker is unable to distinguish multi-potent neural crest progenitor cells from specified glia. An alternative widely used neural crest cell marker is activating enhancer binding protein 2 alpha (AP2α) [17], which like Sox10, is expressed as neural crest cells

delaminate from the neural tube and during their early migration toward the forming DRG.

Analysis of AP2α and Sox10-positive neural crest cells at E9.25 revealed no deficiency of early neural crest cell delamination and migration in the trunk of *Wnt1-Cre; Mib1^{fl/fl}* embryos (n=3/3; Fig. 4a, d). Likewise, at E9.5, there was no difference in the number of AP2α-positive neural crest cells between wildtype and *Wnt1-Cre;*

Mib1^{fl/fl} embryos (n=4/4; Fig. 4b, e). However, there was a notable decrease in the number of Sox10-positive neural crest cells in *Wnt1-Cre; Mib1^{fl/fl}* embryos at this stage (Fig. 4b, e), suggesting a subset of AP2α-positive neural crest cells had begun to downregulate expression of Sox10 during migration between E9.25 and E9.5. To determine the fate of these AP2α-positive, Sox10-negative neural crest cells, trunk neural crest cell development



was analysed slightly later at E9.75. While in wildtype embryos a small proportion of cells had begun to express the neuronal marker *Isl1* with the majority of cells still expressing *Sox10*, in *Wnt1-Cre; Mib1^{fl/fl}* embryos around half of all neural crest cells populating the DRG had begun to express *Isl1*, at the expense of *Sox10* ($n=3/3$; Fig. 4c, f). This is consistent with the premature expression of Trk receptors observed at E10.5. Taken together, this data suggests that initial neural crest cell specification, delamination and migration occurs normally in *Wnt1-Cre; Mib1^{fl/fl}* embryos, however, shortly upon exiting the neural tube, a subset of neural crest cells rapidly lose expression of *Sox10*, and as a consequence differentiate into neurons, without a change in the overall number of neural crest progenitors populating the DRG. This also implies that neuroglial fate choice of neural crest cells forming the DRG may occur earlier in development than previously reported [4, 8, 18].

Neural crest cells biased toward the sensory lineage form correctly in the absence of *Mib1*

While our previous results suggest that neural crest cell formation and delamination is not affected in the absence of *Mib1* they did not address a potential role for Notch signalling in regulating the bias of neural crest cells toward the sensory versus sympathetic lineage. The pro-neural transcription factor *Neurogenin 2* (*Ngn2*) is required to specify neural crest cells toward the sensory lineage and is expressed in neural crest cells as they delaminate and migrate toward the DRG [2, 19]. A similar number of neural crest cells were found to express *Ngn2* in *Wnt1-Cre; Mib1^{fl/fl}* and control embryos at E9.5 (23 somites) and E10.5 ($n=3/3$; Additional file 1: Fig. S2). *Mib1* is therefore dispensible for regulating expression of *Ngn2* and biasing neural crest cells toward the sensory lineage.

Active Notch signalling and neuroglial fate choice occurs immediately as neural crest cells exit the neural tube

Delta-like 1 (DLL1) has previously been characterised as the major Notch ligand directing neuroglial fate choice during DRG development [8]. *Mib1* functions to regulate endocytosis of Notch ligands upon signalling to Notch receptors on adjacent cells, and this endocytosis is essential to enable correct Notch signalling in the receiving cell. Absence of *Mib1* has previously been shown to result in accumulation of Notch ligands at the cell surface [12], and hence this protein accumulation is reflective of cells that are attempting to actively signal to adjacent cells.

Analysis of DLL1 expression during early neural crest cell development, through to stages of DRG condensation, revealed dynamic changes in DLL1 localisation in

Wnt1-Cre; Mib1^{fl/fl} embryos when compared to wildtype (Fig. 5). At E9.25 (20 somites), co-immunostaining with the neural crest marker p75 neurotrophin receptor (p75) and DLL1 revealed a salt-and-pepper pattern of DLL1 expression within the migrating streams of trunk neural crest cells of wildtype embryos (Fig. 5a). However, in *Wnt1-Cre; Mib1^{fl/fl}* embryos, DLL1 was highly expressed in most migrating neural crest cells, and was particularly abundant in the dorsal-most cells that had just delaminated from the neural tube ($n=3/3$; arrowheads, Fig. 5b). This expression pattern was maintained in wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.5 ($n=3/3$; 23 somites) (Fig. 5c, d). This suggests active DLL1-Notch signalling is occurring between multipotent neural crest progenitors at the earliest stages of neural crest migration post-delamination.

At later stages of neural crest migration and early DRG condensation, co-immunostaining for *Tuj1* and DLL1 revealed minor changes in DLL1 localisation and abundance in the DRG compared to earlier stages of development. At E9.75 (26 somites), premature differentiation of neuronal cells in the ventral-most region of the forming DRG was evident in *Wnt1-Cre; Mib1^{fl/fl}* embryos as shown by increased *Tuj1* immunostaining compared to wildtype ($n=3/3$; Fig. 5e, f). At this stage of development, there was also increased DLL1 immunostaining in the dorsal-most neural crest progenitors populating the DRG in *Wnt1-Cre; Mib1^{fl/fl}* embryos ($n=3/3$; solid arrowheads, Fig. 5f). By E10.5, minimal DLL1 immunostaining was evident in the DRG in both wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos ($n=3/3$; Fig. 5g, h), suggesting cells in the DRG are not actively undergoing DLL1-Notch signalling at this stage of development. This further suggests DLL1-Notch signalling to determine DRG cell neuroglial fate choice is executed at earlier stages of development, primarily during neural crest cell delamination and migration between E9.25 and E9.5.

The accumulation of DLL1 in the neural crest cells of *Wnt1-Cre; Mib1^{fl/fl}* embryos is also due to a specific effect of *Mib1* on the DLL1 ligand, as other ligands such as Jagged 1 (*Jag1*) did not show aberrant localisation or accumulation in *Wnt1-Cre; Mib1^{fl/fl}* embryos ($n=3/3$; Additional file 1: Fig. S3). An accumulation of DLL1 was also observed at the dorsal ridge of the neural tube in *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.75–E10.5 ($n=3/3$; Fig. 5f, h), which may suggest a role for Notch signalling in roof plate formation, however this likely occurs independently of DRG development.

Upon stimulation with ligand, the Notch receptor is cleaved by γ -secretase to generate an intracellular fragment known as the Notch Intracellular Domain (NICD) which translocates to the nucleus and associates with other protein partners, such as RBPJ and modulation

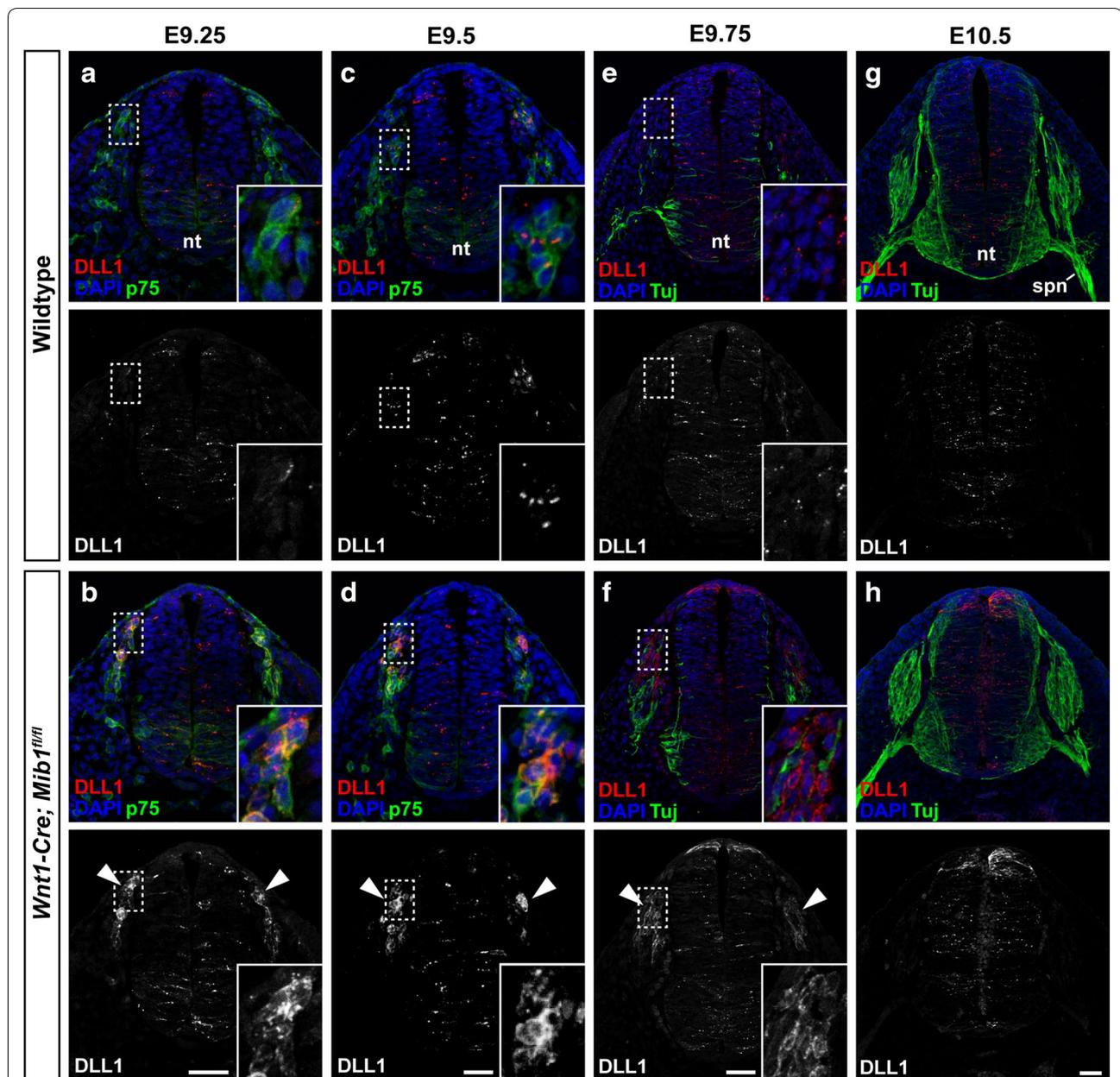
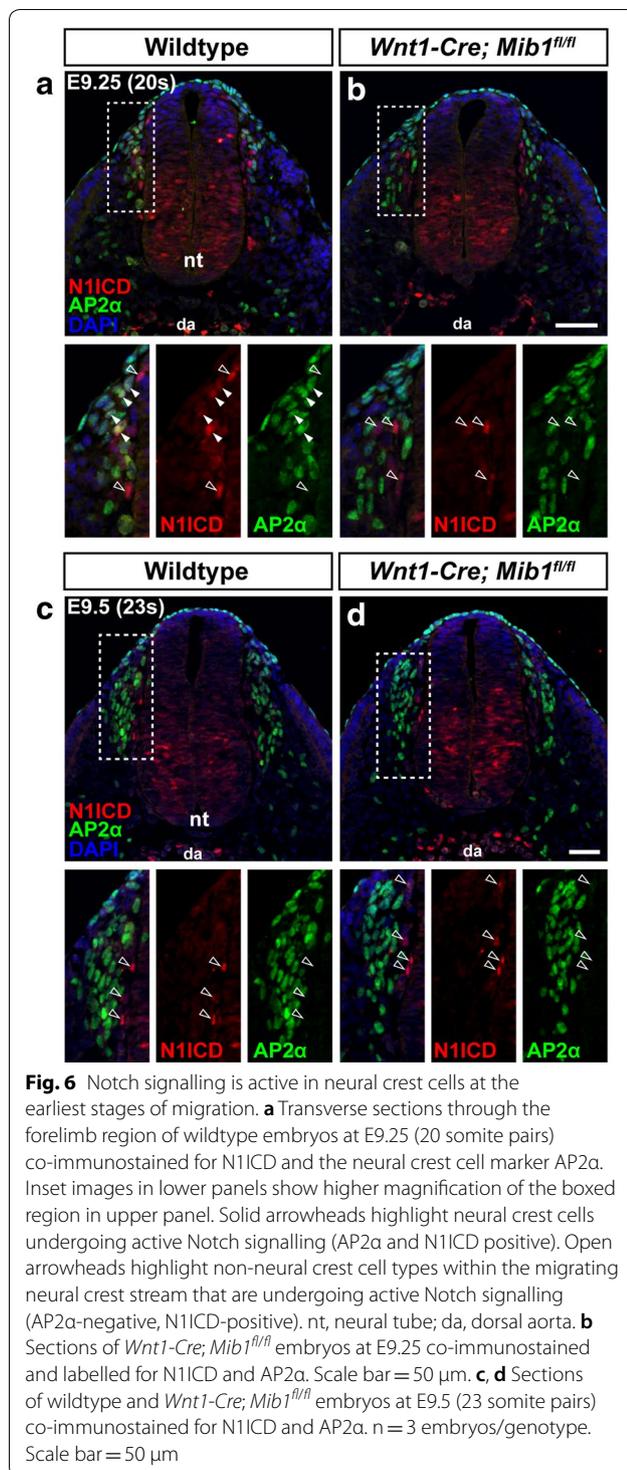


Fig. 5 DLL1 accumulation in *Wnt1-Cre; Mib1^{fl/fl}* embryos reveals timing of DLL1-Notch signalling during early neural crest cell migration. **a** Transverse sections through the forelimb region of wildtype embryos at E9.25 (20 somite pairs) co-immunostained for DLL1 and the neural crest cell marker p75. Lower panel shows DLL1 staining alone for clarity. Inset image is a higher magnification of the boxed region. **b** Sections of *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.25 co-immunostained for DLL1 and p75. Arrowheads indicate regions with increased DLL1 accumulation compared to wildtype. **c, d** Wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.5 (23 somites) co-immunostained for DLL1 and p75. **e, f** Wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.75 (26 somites) co-immunostained for DLL1 and the axonal/neuronal marker TuJ. **g, h** Wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E10.5 co-immunostained for DLL1 and TuJ, n = 3 embryos/genotype. nt neural tube; spn spinal nerve. All scale bars = 50 μm

of Notch signalling by mastermind-like (MAML), to activate target gene transcription. To investigate active Notch signalling during early trunk neural crest cell development, sections were co-immunostained for Notch1 ICD (N1ICD) and AP2α. In wildtype embryos at

E9.25, many of the early migrating (dorsal-most) AP2α-positive neural crest cells were positive for N1ICD (solid arrowheads, Fig. 6a), indicating these cells are undergoing active Notch signalling and have been stimulated by ligand. In *Wnt1-Cre; Mib1^{fl/fl}* embryos however, none



of the AP2α-positive neural crest cells were positive for N1ICD, indicating that in the absence of active ligands, Notch signalling is not induced (n = 3/3; Fig. 6b). This further suggests that the accumulated DLL1 protein present at this stage of development in *Wnt1-Cre; Mib1^{fl/fl}*

embryos is non-functional. As expected, Notch signalling is not abrogated in non-neural crest derivatives in *Wnt1-Cre; Mib1^{fl/fl}* embryos, as evidenced by positive N1ICD staining in the neural tube, endothelial cells of the dorsal aorta (da), and other AP2α-negative cell types present within the migrating neural crest stream (open arrowheads, Fig. 6a, b). Notch signalling was also analysed at E9.5, however none of the neural crest cells in wildtype or *Wnt1-Cre; Mib1^{fl/fl}* embryos were positive for N1ICD (n = 3/3; Fig. 6c, d), suggesting this important DLL1-Notch1 signalling event occurs immediately after neural crest cells have delaminated from the neural tube. Expression of the Notch1 receptor remained unchanged in *Wnt1-Cre; Mib1^{fl/fl}* embryos (Additional file 1: Fig. S4), further suggesting the changes in N1ICD observed were a direct result of inactive ligand-receptor signalling.

Discussion

In this study, we show that removal of *Mib1*, and hence Notch ligand activity, in neural crest-derived tissue causes profound DRG defects. Our findings suggest that a loss of Sox10 expression in a subset of early migrating neural crest cells leads to a reduction of glial cells in the DRG, which is accompanied by neuronal cell death, resulting in severe DRG hypoplasia in later development. While Notch signalling has previously been demonstrated to play an important role in DRG and in particular glial development [5, 6], our study brings into question the timing of this neuroglial fate choice. By taking advantage of DLL1 ligand accumulation in the *Mib1* knockout model, and by demonstrating the timing of N1ICD presence in the nucleus of neural crest cells, our data suggests that active Notch signalling promotes glial cell fate decisions much earlier in DRG development than originally anticipated.

Previous models of neuroglial fate choice in the DRG are largely based on the premise that neural crest cells migrate from the neural tube and coalesce into a condensed DRG structure as an unspecified population of progenitors [3, 4]. Under this model, most nascent neural crest cells are predicted to adopt a neuronal or glial fate after this condensed structure has formed. Another common point of view is that neuronal differentiation occurs before glial specification, where neuronally biased DLL1 expressing cells residing in the core of the DRG signal via Notch to unspecified cells in the DRG periphery to prevent their neuronal differentiation and promote glial specification [3, 8, 20, 21]. A number of our findings instead fit with the notion that Notch signalling occurs between neural crest cells at the earliest stages of neural crest migration prior to coalescing into the nascent DRG structure and before establishment of the core versus periphery. Thus, Notch activity was readily

detected in neural crest cells that had recently delaminated at E9.25 but not in the neural crest cells coalescing within the region of the DRG at any stage examined. Following the absence of Notch signalling in *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.25 we also identified an increase in Isl1 positive neurons in the dorsal regions of the DRG at E9.75. The observation that these ectopic Isl1 positive cells colocalise with Dll1 positive neural crest cells that have attempted to activate Notch during their migration further suggests that these cells aberrantly form neurons. However, in the absence of lineage tracing cells that have activated Notch during early neural crest cell migration it remains possible that the neuronal or glial fate choice could also be influenced at earlier or later stages of development. Indeed, a limitation of our mouse model is that Mib1 has only been removed in neural crest cells as they delaminate which has not allowed us to address any potential roles for Notch activity in premigratory neural crest in the dorsal neural tube at earlier stages of development.

The question of whether neural crest progenitors are fate-restricted even before their exit from the neural tube is a heavily debated topic. Spatial transcriptional profiling has revealed distinct sub-populations of neural crest stem cells in the dorsal neural tube, however their concomitant expression of both pluripotency and lineage markers suggests these populations are open to many fate-choices [22]. Indeed, both historical and more recent genetic lineage tracing studies have concluded that the majority of individual premigratory and migratory neural crest cells are multipotent in vivo [23, 24]. However, other studies suggest that the position within and timing of delamination from the neural tube determines the fate of pre-migratory neural crest cells [25], which is also supported by the findings that Ngn2 expressing cells within the dorsal neural tube are biased toward the sensory lineage [2]. In our present study, N1ICD and Dll1 were not detected in the premigratory neural crest of the dorsal neural tube. We observe active Notch signalling in neural crest cells only after delamination from the neural tube, which supports the notion that DRG-derivatives are not fate-restricted prior to delamination, and that neuroglial fate choice is determined once neural crest cells are migratory.

Previous work has shown that inhibition of Notch signalling (via deletion of *Rbpj*) in the DRG leads to an increase in the number of neurons at the expense of glial cells [9], implying a change of balance in a binary fate-choice decision. In our model, removal of Notch signalling (via deletion of *Mib1*) lead to a decrease in glial cells, without a concomitant overall increase in neuronal cells at E10.5. We did, however, observe an initial premature differentiation of Isl1-positive neuronal cells at E9.75,

and increased numbers of Trk-positive, Isl1-positive co-stained cells at E10.5. Given the number of Isl1-positive neurons remained normal at E10.5, our data suggests the neuronal differentiation/maturation program (Isl1 expression, followed by Trk expression) is executed prematurely in our model. The lack of cell death observed between E9.75 and E10.5 further rules out the possibility that supernumerary neurons are dying. Our data fit with a model in which the number of neurons within the DRG is capped at a fixed number, and that this is reached by E10.5 in *Mib1* conditional knockouts due to premature neuronal differentiation. While the exact nature of this phenomenon remains unclear, the differences between our data and that from *Rbpj* deletion may indicate other roles for *Rbpj* and/or *Mib1* outside of the Notch pathway, or that some Notch activity remains in these models. Notably, Notch has been shown to signal independently of *Rbpj* in some circumstances [26], and *Rbpj* has Notch-independent transcriptional targets [27]. *Mib1* also has roles outside of regulating Notch ligands [28–30], and it is likely that other ubiquitinated targets of *Mib1* are yet to be identified, which may contribute to the phenotype observed.

In addition to inhibiting neuronal differentiation, Notch signalling has also been shown to promote glial cell differentiation in cultured rat neural crest stem cells [7]. Consistent with this idea, we found that Notch activity is necessary to drive expression of the glial markers, Sox10 and FABP7, and to promote gliogenesis. Our data also indicate that one of the primary roles for Notch signalling is to maintain Sox10 expression within a subset of neural crest cells which likely represent glial progenitors. Thus, we show that the number of Sox10 positive cells is reduced prior to an increase in Isl1 positive neuronal precursors. While the mechanisms by which Notch regulates expression of Sox10 remain unknown, it is intriguing that inhibition of Notch in neural crest cells derived from induced pluripotent stem cells leads to Sox10 down regulation [31], and Notch signalling can induce ectopic expression of Sox10 in zebrafish [32]. Taken together with our data this may suggest a direct role for N1ICD in driving Sox10 expression under certain circumstances. Indeed, our own bioinformatic analysis has identified two putative highly conserved RBPj binding sites in intron 1 of the Sox10 locus which may also support this suggestion.

By using *Mib1* deletion in this study our intention was to overcome any redundant roles that the Notch ligands may have in neural crest cells. Functional validation of *Mib1* deletion with antibodies against two Notch ligands found that Dll1, but not Jag1, is expressed in a sub-set of neural crest cells migrating toward the DRG. This expression pattern is in agreement with previous work

implicating a role for Dll1 expressing neural crest cells in myogenesis [33, 34]. Deletion of Dll1 in mice leads to an overlapping phenotype to that identified in our work, including reduced glia and sensory neurons within the DRG [11]. Dll3 also appears to be expressed in a sub-set of neural crest cells within the region of the forming DRG [35], but a role for this ligand in sensory neurogenesis has not specifically been addressed. Whether Dll1 acts as the sole Notch ligand in this fate choice, or if other ligands such as Dll3 are also involved, therefore remains to be explored.

One of the notable defects observed in *Mib1* conditional knockouts in this study was aberrant cell death and depletion of sensory neurons within the DRG after E11.5. A similar temporal progression of neuronal cell death in *Sox10* mouse knockouts has been interpreted by some groups to arise as a secondary defect to deficient glial differentiation [36]. However, *Sox10* has also been shown to have instructive and pro-survival effects in sensory neurons which may suggest that glia are not required for cell survival during early stages of neuronal differentiation [37, 38]. It is also noteworthy that sensory neuron death within the DRG begins at E11.5 in mouse mutants lacking the pro-survival neurotrophin receptor TrkC independent of abnormalities in glia [16]. As we have removed *Mib1* from the precursors of both neurons and glia, and it remains possible that *Mib1* may also regulate other proteins outside of the Notch ligands, our data do not allow us to decipher the mechanisms leading to neuronal cell death at this stage.

An interesting observation in our study was mal-positioning of the dorsal root entry zone, misshapen dorsal neural tube, and aberrant axonal tracts in *Wnt1-Cre; Mib1^{fl/fl}* embryos (Figs. 2, 3, 5). *Wnt1-Cre* drives genetic recombination in the dorsal neural tube prior to neural crest delamination and would be expected to remove *Mib1*, and therefore Notch ligand activity, in this region. Indeed, accumulation of Dll1 in the dorsal neural tube of *Wnt1-Cre; Mib1^{fl/fl}* embryos supports this suggestion. As the N1ICD regulated transcription factor, *Hes1*, has previously been shown to play an important role in roof plate formation downstream of BMP [39], this may suggest that Notch signalling intersects with BMP signalling to control correct development and patterning of the dorsal neural tube.

Conclusions

Taken together, our data define the timing at which Notch ligands signal to Notch receptors to determine the neuronal and glial fate of neural crest cells which will give rise to the DRG. This period at which sensory biased neural crest cells choose their fate is much earlier than previously predicted, occurring at the time, or soon after,

delamination. Defining a novel cell-autonomous role for the ubiquitin ligase *Mib1* in fate specification and formation of the sensory nervous system further highlights the necessity to understand how post translational modifications regulate neural crest cell development [40].

Methods

Mouse lines

All experiments on mice were approved by the SA Pathology and University of South Australia Animal Ethics Committee (Research Ethics License 51-15). Mice were kept in open top conventional cages, on a 12-h light/dark cycle, with free access to food and water. To obtain embryos of defined gestational ages, mice were mated in the evening, and the morning of vaginal plug formation was counted as embryonic day (E) 0.5. Pregnant dams were humanely euthanized at relevant days post vaginal plug detection by CO₂ inhalation and cervical dislocation. To lineage trace neural crest cells and their derivatives, we crossed *Wnt1-Cre* mice [41] to *Z/EG* mice [42]. To delete *Mib1* specifically in neural crest cells, we mated *Mib1^{fl/+}* males carrying a heterozygous *Wnt1-Cre* transgene to *Mib1^{fl/fl}* female mice [43]. At least 3 embryos/age/genotype were analysed for each experiment.

Immunohistochemistry

Embryos were fixed in 4% paraformaldehyde for 2 h or overnight at 4 °C, cryopreserved in 20% sucrose, and embedded in OCT compound for cryosectioning. Sections were blocked in 10% Dako serum-free blocking reagent or 10% goat serum in PBS 0.1% TritonX-100 (with some exceptions, see below), followed by incubation in primary antibody for 2 h at room temperature or overnight at 4 °C. Fluorescent Alexafluor-conjugated secondary antibodies were incubated for 1 h at room temperature. Sections were mounted in Prolong Diamond antifade with 4',6-diamidino-2-phenylindole (DAPI). The following primary antibodies were used: Chicken anti-EGFP, 1:1000 (Abcam ab13970); mouse anti-Tuj1, 1:750 (Sigma-Aldrich T5076); goat anti-*Sox10*, 1:100 (Santa Cruz sc-17342); goat anti-*Sox10*, 1:100 (R&D Systems AF2864); mouse anti-*Isl1*, 1:50 (DSHB 40.3A4); rabbit anti-cleaved Caspase-3, 1:500 (Cell Signaling Technologies 9661); rabbit anti-FABP7, 1:200 (Cell Signaling Technologies 13347); goat anti-TrkA, 1:50 (R&D Systems AF1056); goat anti-TrkB, 1:200 (R&D Systems AF1494); goat anti-TrkC, 1:200 (R&D Systems AF1404); mouse anti-AP2 α , 1:20, goat serum block (DSHB 3B5); sheep anti-DLL1, 1:200 (R&D Systems AF5026); rabbit anti-p75, 1:250 (Abcam 52987); goat anti-Jag1, 1:200 (R&D Systems AF599); rabbit anti-N1ICD, 1:100 (Cell Signaling Technologies 4147); sheep anti-Notch1, 1:200 (R&D Systems AF5267).

Antigen retrieval

Immunostaining for AP2 α and N1ICD required antigen retrieval [44]. Cryosections were incubated in 10 mM Sodium Citrate pH6.0 at 90 °C for 20 min, and then cooled to room temperature before proceeding with immunohistochemistry procedure as above.

Tyramide amplification

Immunostaining for N1ICD required tyramide signal amplification, performed using Invitrogen Alexa Fluor 555 Tyramide SuperBoost Kit (B40923). Following primary antibody incubation, sections were incubated with goat anti-rabbit horse radish peroxidase (HRP), then tyramide signal amplification performed for 30 min, following manufacturer's instructions.

TUNEL staining

Following immunohistochemical staining, sections were stained using Roche In Situ Cell Death Detection Kit TMR red following manufacturer's instructions for 1 h at 37 °C.

In situ hybridisation

Section in situ hybridisation was performed as described [45]. Riboprobes were transcribed from plasmids containing the cDNA sequence for *Ngn2* [46].

Quantitation of cell numbers

Transverse sections spanning the anterior to posterior limits of 2–3 DRG were collected over the forelimbs of E9.25–11.5 embryos. The total number of Sox10 positive, Isl1 positive, TUNEL positive, Casp3 positive, Trk positive or AP2 α positive cells were manually counted from 20 \times images from each section spanning the limits of each DRG. Approximately 15–20 sections each from at least 3 embryos per age/genotype were analysed. Data are represented as the number of antigen-positive cells per 12 μ m section per DRG. All data are presented as mean \pm SEM and analysed using Student's *t* test. In all studies a *p* value of < 0.05 was considered to be statistically significant.

Additional file

Additional file 1: Fig. S1. Glial cell maturation is inhibited in the DRG of *Wnt1-Cre; Mib1^{fl/fl}* mice. **(A)** Transverse sections of wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E11.5 co-immunostained for the axonal marker Tuj1 and the glial marker FABP7. **(B)** FABP7 staining shown alone for clarity. Scale bar = 50 μ m. **(C)** Transverse sections of wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E13.5 co-immunostained for Tuj1 and FABP7. **(D)** FABP7 staining shown alone for clarity. n = 3 embryos/genotype. nt, neural tube; spn, spinal nerve. Scale bar = 100 μ m. **Fig. S2.** Neural crest specification toward the sensory lineage is conserved in *Wnt1-Cre; Mib1^{fl/fl}* mice. In situ hybridisation for *Ngn2* on transverse sections of wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.5 and E10.5. n = 3 embryos/genotype. nt, neural

tube. Scale bar = 50 μ m. **Fig. S3.** The Notch ligand Jag1 remains unchanged in neural crest upon loss of Mib1. Transverse sections of wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.25 co-immunostained for the neural crest cell marker p75 and Jag1. n = 3 embryos/genotype. nt, neural tube. Scale bar = 50 μ m. **Fig. S4.** Total Notch1 receptor levels remain unchanged in neural crest upon loss of Mib1. Transverse sections of wildtype and *Wnt1-Cre; Mib1^{fl/fl}* embryos at E9.5 co-immunostained for the neural crest cell marker p75 and Notch1. n = 3 embryos/genotype. nt, neural tube. Scale bar = 50 μ m.

Abbreviations

DRG: dorsal root ganglia; Mib1: Mindbomb 1; DLL1: Delta-like 1; RBPJ: recombination signal binding protein for immunoglobulin kappa J region; Sox10: SRY-related HMG-box 10; N1ICD: Notch1 intracellular domain; E: embryonic day; EGFP: enhanced green fluorescent protein; Tuj1: neuron specific class-III b tubulin; Isl1: islet 1; TUNEL: terminal deoxynucleotidyl transferase dUTP nick end labelling; Casp3: cleaved caspase 3; AP2a: activating enhancer binding protein 2 alpha; Ngn2: Neurogenin 2; p75: p75 neurotrophin receptor; Jag1: Jagged 1; MAML: modulation of Notch signalling by mastermind-like; da: dorsal aorta; FABP7: fatty acid binding protein 7; PBS: phosphate buffered saline; DAPI: 4',6-diamidino-2-phenylindole; HRP: horse radish peroxidase.

Authors' contributions

SW performed all experiments. SW and QS analysed and interpreted all data and wrote the manuscript. Both authors read and approved the final manuscript.

Acknowledgements

We thank Young-Yun Kong of the Biological Sciences Department, Seoul National University, South Korea, for providing *Mib1^{fl/fl}* mice. We also thank Xiangjun Xu for genotyping the mice and embryos in this study.

Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

All data generated or analysed during this study are included in this published article and its supplementary information files.

Consent for publication

Not applicable.

Ethics approval and consent to participate

All experiments on mice were approved by the SA Pathology and University of South Australia Animal Ethics Committee (Research Ethics License 51-15).

Funding

SW was supported by a Royal Adelaide Hospital Mary Overton Postdoctoral Fellowship. QS was supported by a National Heart Foundation Future Leader Fellowship. Research funding was supported by a NHMRC Project grant to QS (APP106958).

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 3 December 2018 Accepted: 15 April 2019

Published online: 29 April 2019

References

- Hanani M. Satellite glial cells in sensory ganglia: from form to function. *Brain Res Brain Res Rev.* 2005;48(3):457–76.
- Zirlinger M, Lo L, McMahon J, McMahon AP, Anderson DJ. Transient expression of the bHLH factor neurogenin-2 marks a subpopulation of

- neural crest cells biased for a sensory but not a neuronal fate. *Proc Natl Acad Sci USA*. 2002;99(12):8084–9.
3. Jacob C. Transcriptional control of neural crest specification into peripheral glia. *Glia*. 2015;63(11):1883–96.
 4. Marmigere F, Ernfor P. Specification and connectivity of neuronal subtypes in the sensory lineage. *Nat Rev Neurosci*. 2007;8(2):114–27.
 5. Cornell RA, Eisen JS. Notch in the pathway: the roles of Notch signaling in neural crest development. *Semin Cell Dev Biol*. 2005;16(6):663–72.
 6. Louvi A, Artavanis-Tsakonas S. Notch signalling in vertebrate neural development. *Nat Rev Neurosci*. 2006;7(2):93–102.
 7. Morrison SJ, Perez SE, Qiao Z, Verdi JM, Hicks C, Weinmaster G, Anderson DJ. Transient Notch activation initiates an irreversible switch from neurogenesis to gliogenesis by neural crest stem cells. *Cell*. 2000;101(5):499–510.
 8. Wakamatsu Y, Maynard TM, Weston JA. Fate determination of neural crest cells by NOTCH-mediated lateral inhibition and asymmetrical cell division during gliogenesis. *Development*. 2000;127(13):2811–21.
 9. Hu ZL, Shi M, Huang Y, Zheng MH, Pei Z, Chen JY, Han H, Ding YQ. The role of the transcription factor Rbpj in the development of dorsal root ganglia. *Neural Dev*. 2011;6:14.
 10. Taylor MK, Yeager K, Morrison SJ. Physiological Notch signaling promotes gliogenesis in the developing peripheral and central nervous systems. *Development*. 2007;134(13):2435–47.
 11. De Bellard ME, Ching W, Gossler A, Bronner-Fraser M. Disruption of segmental neural crest migration and ephrin expression in Delta-1 null mice. *Dev Biol*. 2002;249(1):121–30.
 12. Kang K, Lee D, Hong S, Park SG, Song MR. The E3 ligase Mind bomb-1 (Mib1) modulates Delta-Notch signaling to control neurogenesis and gliogenesis in the developing spinal cord. *J Biol Chem*. 2013;288(4):2580–92.
 13. Britsch S, Goerich DE, Riethmacher D, Peirano RI, Rossner M, Nave KA, Birchmeier C, Wegner M. The transcription factor Sox10 is a key regulator of peripheral glial development. *Genes Dev*. 2001;15(1):66–78.
 14. Kim J, Lo L, Dormand E, Anderson DJ. SOX10 maintains multipotency and inhibits neuronal differentiation of neural crest stem cells. *Neuron*. 2003;38(1):17–31.
 15. Le Douarin N, Kalcheim C. The neural crest. 2nd ed. Cambridge: Cambridge University Press; 1999.
 16. White FA, Silos-Santiago I, Molliver DC, Nishimura M, Phillips H, Barbacid M, Snider WD. Synchronous onset of NGF and TrkA survival dependence in developing dorsal root ganglia. *J Neurosci*. 1996;16(15):4662–72.
 17. Mitchell PJ, Timmons PM, Hebert JM, Rigby PW, Tjian R. Transcription factor AP-2 is expressed in neural crest cell lineages during mouse embryogenesis. *Genes Dev*. 1991;5(1):105–19.
 18. Abe M, Ruest LB, Clouthier DE. Fate of cranial neural crest cells during craniofacial development in endothelin-A receptor-deficient mice. *Int J Dev Biol*. 2007;51(2):97–105.
 19. Ma Q, Fode C, Guillemot F, Anderson DJ. Neurogenin1 and neurogenin2 control two distinct waves of neurogenesis in developing dorsal root ganglia. *Genes Dev*. 1999;13(13):1717–28.
 20. Morrison SJ. Neuronal potential and lineage determination by neural stem cells. *Curr Opin Cell Biol*. 2001;13(6):666–72.
 21. Carr VM, Simpson SB Jr. Proliferative and degenerative events in the early development of chick dorsal root ganglia. II. Responses to altered peripheral fields. *J Comp Neurol*. 1978;182(4):741–55.
 22. Lignell A, Kerosuo L, Streichan SJ, Cai L, Bronner ME. Identification of a neural crest stem cell niche by spatial genomic analysis. *Nat Commun*. 2017;8(1):1830.
 23. Bronner-Fraser M, Fraser SE. Cell lineage analysis reveals multipotency of some avian neural crest cells. *Nature*. 1988;335(6186):161–4.
 24. Baggiolini A, Varum S, Mateos JM, Bettosini D, John N, Bonalli M, Ziegler U, Dimou L, Clevers H, Furrer R, et al. Premigratory and migratory neural crest cells are multipotent in vivo. *Cell Stem Cell*. 2015;16(3):314–22.
 25. Krispin S, Nitzan E, Kassem Y, Kalcheim C. Evidence for a dynamic spatiotemporal fate map and early fate restrictions of premigratory avian neural crest. *Development*. 2010;137(4):585–95.
 26. Turkoz M, Townsend RR, Kopan R. The Notch intracellular domain has an RBPj-independent role during mouse hair follicular development. *J Invest Dermatol*. 2016;136(6):1106–15.
 27. Diaz-Trelles R, Scimia MC, Bushway P, Tran D, Monosov A, Monosov E, Peterson K, Rentschler S, Cabrales P, Ruiz-Lozano P, et al. Notch-independent RBPJ controls angiogenesis in the adult heart. *Nat Commun*. 2016;7:12088.
 28. Mizoguchi T, Ikeda S, Watanabe S, Sugawara M, Itoh M. Mib1 contributes to persistent directional cell migration by regulating the Ctnnd1-Rac1 pathway. *Proc Natl Acad Sci USA*. 2017;114(44):E9280–9.
 29. Kwon DY, Dimitriadis M, Terzic B, Cable C, Hart AC, Chitnis A, Fischbeck KH, Burnett BG. The E3 ubiquitin ligase mind bomb 1 ubiquitinates and promotes the degradation of survival of motor neuron protein. *Mol Biol Cell*. 2013;24(12):1863–71.
 30. Matsuda M, Rand K, Palardy G, Shimizu N, Ikeda H, Dalle Nogare D, Itoh M, Chitnis AB. Epb4115 competes with Delta as a substrate for Mib1 to coordinate specification and differentiation of neurons. *Development*. 2016;143(17):3085–96.
 31. Noisa P, Lund C, Kanduri K, Lund R, Lahdesmaki H, Lahesmaa R, Lundin K, Chocheuwattanalert H, Otonkoski T, Tuuri T, et al. Notch signaling regulates the differentiation of neural crest from human pluripotent stem cells. *J Cell Sci*. 2014;127(Pt 9):2083–94.
 32. Dutton JR, Antonellis A, Carney TJ, Rodrigues FS, Pavan WJ, Ward A, Kersh RN. An evolutionarily conserved intronic region controls the spatiotemporal expression of the transcription factor Sox10. *BMC Dev Biol*. 2008;8:105.
 33. Nandagopal N, Santat LA, LeBon L, Sprinzak D, Bronner ME, Elowitz MB. Dynamic ligand discrimination in the Notch signaling pathway. *Cell*. 2018;172(4):869–880 e819.
 34. Rios AC, Serralbo O, Salgado D, Marcelle C. Neural crest regulates myogenesis through the transient activation of NOTCH. *Nature*. 2011;473(7348):532–5.
 35. Dunwoodie SL, Henrique D, Harrison SM, Beddington RS. Mouse Dll3: a novel divergent Delta gene which may complement the function of other Delta homologues during early pattern formation in the mouse embryo. *Development*. 1997;124(16):3065–76.
 36. Sonnenberg-Riethmacher E, Miehle M, Stolt CC, Goerich DE, Wegner M, Riethmacher D. Development and degeneration of dorsal root ganglia in the absence of the HMG-domain transcription factor Sox10. *Mech Dev*. 2001;109(2):253–65.
 37. Carney TJ, Dutton KA, Greenhill E, Delfino-Machin M, Dufourcq P, Blader P, Kersh RN. A direct role for Sox10 in specification of neural crest-derived sensory neurons. *Development*. 2006;133(23):4619–30.
 38. Delfino-Machin M, Madelaine R, Busolin G, Nikaïdo M, Colanesi S, Camargo-Sosa K, Law EW, Toppo S, Blader P, Tiso N, et al. Sox10 contributes to the balance of fate choice in dorsal root ganglion progenitors. *PLoS ONE*. 2017;12(3):e0172947.
 39. Nitzan E, Avraham O, Kahane N, Ofek S, Kumar D, Kalcheim C. Dynamics of BMP and Hes1/Hairy1 signaling in the dorsal neural tube underlies the transition from neural crest to definitive roof plate. *BMC Biol*. 2016;14:23.
 40. Wiszniak S, Lumb R, Kabbara S, Scherer M, Schwarz Q. Li-gazing at the crest: modulation of the neural crest by the ubiquitin pathway. *Int J Biochem Cell Biol*. 2013;45:1087–91.
 41. Danielian PS, Muccino D, Rowitch DH, Michael SK, McMahon AP. Modification of gene activity in mouse embryos in utero by a tamoxifen-inducible form of Cre recombinase. *Curr Biol*. 1998;8(24):1323–6.
 42. Novak A, Guo C, Yang W, Nagy A, Lobe CG. Z/EG, a double reporter mouse line that expresses enhanced green fluorescent protein upon Cre-mediated excision. *Genesis*. 2000;28(3–4):147–55.
 43. Koo BK, Lim HS, Song R, Yoon MJ, Yoon KJ, Moon JS, Kim YW, Kwon MC, Yoo KW, Kong MP, et al. Mind bomb 1 is essential for generating functional Notch ligands to activate Notch. *Development*. 2005;132(15):3459–70.
 44. Del Monte G, Grego-Bessa J, Gonzalez-Rajal A, Bolos V, De La Pompa JL. Monitoring Notch1 activity in development: evidence for a feedback regulatory loop. *Dev Dyn*. 2007;236(9):2594–614.
 45. Schwarz Q, Gu C, Fujisawa H, Sabelko K, Gertsenstein M, Nagy A, Taniguchi M, Kolodkin AL, Ginty DD, Shima DT, et al. Vascular endothelial growth factor controls neuronal migration and cooperates with Sema3A to pattern distinct compartments of the facial nerve. *Genes Dev*. 2004;18(22):2822–34.
 46. Cau E, Gradwohl G, Fode C, Guillemot F. Mash1 activates a cascade of bHLH regulators in olfactory neuron progenitors. *Development*. 1997;124(8):1611–21.